

Potential Implications of Organoids in Dental Research

Deepti Sharma^{1*}, George Koshy¹, Merin S George¹, Vishal Kumar Sharma²

1. Department of Oral and Maxillofacial Pathology, Christian Dental College & Hospital, Ludhiana, Punjab, India

2. Department of Orthodontics, Desh Bhagat Dental College, Mandi Gobindgarh, Punjab, India

Article Info

ABSTRACT

Article type:

Review Article

Article History:

Received: 21 Jul 2024

Accepted: 19 Feb 2025

Published: 20 Aug 2025

Organoids are among the most significant advancements in the past few years that have revolutionized the medical and dental research. These experimental organs have been used to study the developmental processes, pathophysiology, genetic mutations, and drug effects. The organoid technology is an important adjunct to two-dimensional culture methods and animal models. Stem cells are grown in specific environments to develop the organoids, thus, simulating the complexity of the in vivo environment. Oral organoids also recapitulate the architecture and functions of in vivo organs and tissues in the oral and maxillofacial region. Organoids are a promising tool to manipulate human dental stem cell biology for comprehending tooth-regenerative approaches, infectious diseases, genetic anomalies, oral cancer, and drug discovery.

* Corresponding author:

Department of Oral and Maxillofacial Pathology, Christian Dental College and Hospital, Ludhiana, India

Keywords: Organoids; Stem Cells; Regenerative Medicine; Molecular Targeted Therapy; Tissue Engineering

Email: deepti_dentist@yahoo.co.in

➤ **Cite this article as:** Sharma D, Koshy G, George MS, Sharma VK. Potential implications of Organoids in Dental Research. *Front Dent.* 2025;22:33. <http://doi.org/10.18502/fid.v22i33.19561>

INTRODUCTION

Over the decades, animal model systems, cell lines, two-dimensional tissue cultures, and patient-derived xenograft models have laid the foundation of biomedical research to understand the development process, disease pathogenesis, cellular interactions, signaling pathways, drug discovery, and targeted therapies. But these models could never completely simulate or substitute the in vivo environment [1]. A unique technology, named organoids, has slowly paved the way to revolutionize the 21st century research. Organoids refer to in vitro cells presented in a three-dimensional setup to form clusters of regenerative cells with the ability to recreate the architecture and physiology of human organs by differentiation and self-organization into

various cell forms [2]. This research tool is a human in vitro three-dimensional cell culture resembling greater extent actual human organs anatomically, physiologically, and histologically [3]. This technology has facilitated the understanding of development, course, and nature of disorders, targeting personalized treatment approaches, and has opened new avenues for human disease modeling, tissue engineering, drug development, diagnosis, and regenerative medicine [4]. This manuscript aimed to highlight the use of organoids in therapeutic sciences and regeneration, and its perspectives in dental research.

METHODS

Organoid formation: key aspects

This technology was highlighted first in 2009,

when self-organizing intestinal crypt-villus units were produced utilizing adult stem cells [3]. In the past few years, organoids have been developed for different organs including the brain, liver, colon, pancreas, stomach, retina, thyroid, and reproductive organs[5]. Organoids better simulate the cellular environments, tissue structure, and development process in vivo than the conventional two-dimensional cell culture as these are derived from primary tissue or stem cells mainly from pluripotent stem cells (PSCs) or induced pluripotent cells (iPSCs) and adult stem cells; thus, mimicking human development or organ regeneration in vitro[6]. The quality and capacity of the foundation cells are the key for well-functioning complex and intact organoid models. They can also be generated from patient-derived iPSCs for building organoid replicas of infectious diseases, genetic disorders, degenerative diseases, and cancer, which can be used to create disease-specific research models to uncover intricate details of pathology [7].

Organoids can be derived from all three germ layers and simulate healthy as well as diseased tissues [6]. Stem cell-containing tissue samples are propagated in an appropriate extracellular matrix (Matrigel® is commonly used for this purpose). The cells are cultured in presence of cell culture media containing specific growth factors under controlled conditions that mimic the in vivo environment [8]. Cells further expand rapidly in culture while remaining genetically stable, and self-organize into 3D organoid structures incorporating cellular complexity of the modeled tissue. They can be passaged, and maintained indefinitely [9]. There are definite benefits for human organoid cultures over animal models such as faster and more robust outcomes, providing more quantity of material to work with than animal models, easy handling, feasibility of genetic manipulation, and personalization [1].

Organoids in the medical field:

Organoids can be generated through genetic engineering of human stem cells or from patient biopsy samples to study the disease pattern, and are becoming an asset in terms of modern medicine with their role in testing the drug efficacy in different controlled

microculture environments [10]. Organoids are providing groundbreaking evidence in the study of diseases and development of effective treatments as follows:

- Human adult stem cell-derived organoids in the study of infection biology [11].
- Organoids have also been used to co-cultivate human epithelia with bacteria (for example, *Helicobacter pylori*) and with protozoan parasites (for example, *Cryptosporidium*) [1].
- Organoids for cystic fibrosis [12].
- Human brain development and Zika virus [13].
- Human organoids and severe acute respiratory syndrome-coronavirus [14].

Cancer biology has always been intriguing, and a lot of research is being done by the scientists all over the world. Organoids are revolutionizing the oncology field as well. Preclinical cancer models, animal experiments, and tissue cultures are essential to investigate tumor heterogeneity, genetics, epigenetics, and immunology [15]. Organoid culture models can be harnessed to study normal and neoplastic tissues using similar culture techniques, and are excellent tools to study tumorigenesis and cancer progression. Such samples can then be used to study cancer cell biology, their interactions with the environment, impacts of growth factors on cancer cells, signaling mechanisms, new drug molecules against potential targets, omics analysis, and genetic manipulation [16]. Various patient samples from colon, brain, liver, breast, bladder, prostate, pancreas, stomach, esophageal, endometrial, and lung cancers have readily been cultured as cancer organoid models [17]. Recently, an analysis of drug responses in patients and in their matched cancer organoids led to the conclusion that a drug with no anti-tumor activity in the organoids had the same efficacy and response in the matched patients. Moreover, pathogenic genes and mutations can be directly tested in organoids derived from cells isolated from healthy donors; thus, enabling human genetic studies in controlled genetic backgrounds [10].

Organoids in dentistry:

The organoid technology is being explored in dentistry to recapitulate the oral developmental

process, diseases, genetic disorders, and regeneration of dental tissues (Fig.1).

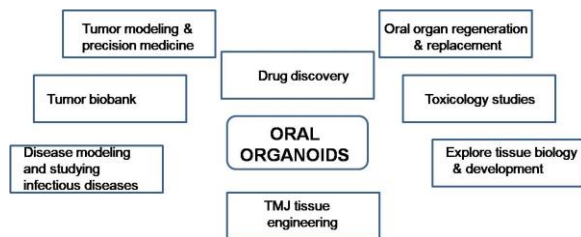


Fig 1. Applications of organoids in dentistry

Many experiments have been previously carried out combining biology and

engineering principles using innovative biomaterials to achieve dental tissue regeneration, by impregnating and growing stem cells on a scaffold with the bioregulatory functions of odontogenic differentiation [18]. Organoids could be apathbreaking advancement towards regenerating the whole tooth structure and other oral tissues. Technological advancements have been made in development of organoids of the tooth, salivary glands, taste buds, tongue, pulp-dentin complex, and also oral cancer tissue [19-29] (Table1).

Table 1. Oral organoids and their applications in dentistry

Number	Organoid derived	Source	Function
1	Tooth germ organoids [19]	Adult human dental pulp cells	To study basic tooth developmental mechanisms
2	Epithelial organoids [20] (human tooth)	Stem cell from dental follicle of 3rd molar	To comprehend human tooth epithelial stem cell biology and epithelium-mesenchyme interaction.
3	Enamel organoid [21]	iPSC-derived ameloblasts	Attempt towards regenerative dentistry and treatment prospects for enamel disorders.
4	Tooth-bud organoids [18]	Human DPSCs, porcine dental epithelial cells	To understand tooth regeneration strategies
5	Pulp organoid[22]	Human dental pulp cells and endothelial cells	For toxicity screening of dental materials on dental pulp cells and tissue
6	Dentin-pulp organoids [23]	Human dental-pulp stem cells	For drug efficacy testing
7	Dental follicle cell organoid [24]	Human dental follicle cells	To comprehend functional PDL engineering
8	Salivary glands [25]	Adult human salivary gland stem/progenitor cells	For treating radiation-induced xerostomia As a novel therapeutic strategy to treat salivary gland dysfunction
9	Taste buds[26]	Source -Lgr5+ stem cells from the circumvallate papillae(murine) Bmi1-positive cells	To explore taste signaling mechanisms, disease modeling and taste tissue regeneration.
10	Lingual epithelium [27]	isolated from the basal layer of the lingual epithelium(mice)	To understand lingual epithelium, lingual regeneration and carcinogenesis.
11	Human mucosal organoids[28]	Primary oral squamous cell carcinoma cells Primary normal keratinocytes	To expand the repertoire of head and neck squamous cell carcinoma drugs and treatment personalization.
12	Esophageal head and neck and oral cancer organoid[29]	Tumoral tissue	Predicting drug sensitivity and potential therapy resistance mechanisms in a moderate-to-high throughput manner, esophageal cancer patient-derived organoids (PDO) are highly translatable in personalized medicine.

Tooth is an easily accessible and non-invasive source of stem cells with self-renewal capabilities and the ability to differentiate into three different lineages. Different stem cell populations can be isolated from dental tissues such as dental pulp stem cells (DPSCs), periodontal ligament (PDL) stem cells, dental follicle progenitor stem cells, stem cells from human exfoliated deciduous teeth, and stem cells from apical papilla [30].

The experiments to characterize phenotype and biological function of human tooth epithelial stem cells are being conducted on the same ground as stem cells of mesenchymal origin so that epithelial-mesenchymal interactions leading to odontogenic differentiation can be simulated in vitro. Recent research has substantiated that the epithelial cell rests of Malassez, present in the dental follicle or PDL around the root can be the potential source of epithelial stem cells [31]. Thus, co-culturing of epithelial cell rests of Malassez-derived stem cells with DPSCs could induce ameloblast differentiation. Hence, dental organoid technology can be used to treat various dental infections, and developmental and genetic anomalies like amelogenesis / dentinogenesis imperfect [19]. Future research needs to emphasize on translating these in vitro findings to clinical practice, paving the way for realization of dental tissue regeneration and less reliance on dental restorations and prosthetics [32].

Organoids from dental pulp mesenchymal stem cells possess strong immunomodulatory properties, and the capacity for angiogenesis, growth factor expression, and formation of vascular pulp tissue [33]. Also, regeneration of PDL with dental follicle cells using an organoid model has been reported, which can be useful to treat PDL defects and periodontitis [34]. The underlying mechanism of taste bud genesis and taste function can be explored by taste bud organoids; thus, paving the way for taste bud regeneration, which can be helpful in oral cancer patients with taste dysfunction. Maxillofacial cartilage regeneration has also been attempted to develop cartilage organoids with superior chondrogenic activity and temporomandibular joint tissue engineering

[35]. Lingual epithelial renewal and regeneration mechanisms have also been studied using organoid models as the incidence of tongue cancer is increasing with significant morbidity [26].

Research has reported the development of bioengineered tooth using an organoid tooth germ model. The three-dimensional salivary gland organoids have been successfully structured and have proven to be quite helpful in understanding the mechanisms to restore normal salivary gland function, treating salivary gland pathologies and xerostomia [25]. Lassche et al. [35] used drug screening organoids treated with tropomyosin receptor kinase-inhibitors to predict in vivo treatment response while preserving tumor genotype and phenotype in a case of ETV6-NTRK3 gene fusion-positive secretory carcinoma of the salivary glands; thus, illustrating the advances made in precision oncology and targeted therapy [36]. Organoids have also revolutionized the investigations of infectious diseases as stem cell-derived organoids from oral mucosal tissue have been developed for viral infections caused by human papillomavirus, herpes simplex virus, etc. Many characteristics of in vivo diseases can be reduplicated by organoid models, providing new insights for disease pathogenesis and host-microbiome interactions [37].

Three-dimensional technology can also be harnessed to understand the influence of tumor microenvironment on oral carcinogenesis and tumor progression [37]. With regard to head and neck squamous cell carcinoma (HNSCC), there is a dire need to have reliable models to explore resistance-related mechanisms, guide treatment decisions, predict therapy outcome, and adopt personalized approaches [38]. Driehuis et al. [27] stated that HNSCC-derived organoids reconstructed the genetic, functional, and histological characteristics of original tumors; they could predict patient response, and widen the horizon of HNSCC drugs. Although the technology has its own specific limitations like lacking the holistic representation of tumor microenvironment, standardized protocols, and cost restraints, yet an

exponential surge has been observed in the recent years for utilizing organoids as models of biological processes and diverse pathophysiological states. A lot more can be explored in the field of oral oncology [39].

CONCLUSION

The promising role of organoid technology in strengthening the existing model systems, translational research, and drug discovery is widely appreciated. This technology has a lot of potential in dentistry but has to be judiciously explored considering its strengths and limitations.

ACKNOWLEDGEMENT

We acknowledge the support provided by Department of Oral & Maxillofacial Pathology, Christian Dental College.

CONFLICT OF INTEREST STATEMENT

Authors declare no conflict of interest.

REFERENCES

- Kim J, Koo BK, Knoblich JA. Human organoids: model systems for human biology and medicine. *Nat Rev Mol Cell Biol.* 2020 Oct;21(10):571-84.
- Simian M, Bissell MJ. Organoids: A historical perspective of thinking in three dimensions. *J Cell Biol.* 2017 Jan;216(1):31-40.
- Corrò C, Novellademunt L, Li VSW. A brief history of organoids. *Am J Physiol Cell Physiol.* 2020 Jul 1;319(1):C151-65.
- Schutgens F, Clevers H. Human organoids: Tools for understanding biology and treating diseases. *Annu Rev Pathol.* 2020 Jan;15:211-34.
- Rauth S, Karmakar S, Batra SK, Ponnusamy MP. Recent advances in organoid development and applications in disease modeling. *BiochimBiophys Acta Rev Cancer.* 2021 Apr;1875(2):188527.
- Clevers H. Modeling development and disease with organoids. *Cell.* 2016 Jun;165(7):1586-97.
- Shariati L, Esmaeili Y, Haghjooy Javanmard S, Bidram E, Amini A. Organoid technology: Current standing and future perspectives. *Stem Cells.* 2021 Dec;39(12):1625-49.
- Lehmann R, Lee CM, Shugart EC, Benedetti M, Charo RA, Gartner Z, et al. Human organoids: a new dimension in cell biology. *Mol Biol Cell.* 2019 May;30(10):1129-37.
- Shankaran A, Prasad K, Chaudhari S, Brand A, Satyamoorthy K. Advances in development and application of human organoids. *3 Biotech.* 2021 Jun;11(6):257.
- Aguilar C, Alves da Silva M, Saraiva M, Neyazi M, Olsson IAS, Bartfeld S. Organoids as host models for infection biology - a review of methods. *Exp Mol Med.* 2021 Oct;53(10):1471-82.
- de Poel E, Lefferts JW, Beekman JM. Intestinal organoids for cystic fibrosis research. *J Cyst Fibros.* 2020 Mar;19 Suppl 1:S60-4.
- Su X, Yue P, Kong J, Xu X, Zhang Y, Cao W, et al. Human brain organoids as an in vitro model system of viral infectious diseases. *Front Immunol.* 2022 Jan;12:792316.
- Kim J, Koo BK, Clevers H. Organoid studies in COVID-19 research. *Int J Stem Cells.* 2022 Feb;15(1):3-13.
- Hoarau-Véhot J, Rafii A, Touboul C, Pasquier J. Halfway between 2D and animal models: Are 3D cultures the ideal tool to study cancer-microenvironment interactions? *Int J Mol Sci.* 2018 Jan;19(1):181.
- Xu H, Jiao D, Liu A, Wu K. Tumor organoids: applications in cancer modeling and potentials in precision medicine. *J Hematol Oncol.* 2022 May;15(1):58.
- Verduin M, Hoeben A, De Ruyscher D, Vooijs M. Patient-derived cancer organoids as predictors of treatment response. *Front Oncol.* 2021 Mar;11:641980.
- Kilic Bektas C, Zhang W, Mao Y, Wu X, Kohn J, Yelick PC. Self-assembled hydrogel microparticle-based tooth-germ organoids. *Bioengineering (Basel).* 2022 May;9(5):215.
- Rosowski J, Bräunig J, Amler AK, Strietzel FP, Lauster R, Rosowski M. Emulating the early phases of human tooth development in vitro. *Sci Rep.* 2019 May;9(1):7057.
- Hemeryck L, Hermans F, Chappell J, Kobayashi H, Lambrechts D, Lambrechts I, et al. Organoids from human tooth showing epithelial stemness phenotype and differentiation potential. *Cell Mol Life Sci.* 2022 Feb;79(3):153.
- Alghadeer A, Hanson-Drury S, Ehnes D, Zhao YT, Patni AP, O'Day D, Spurrell CH, Gogate AA, Phal A, Zhang H, Devi A. Human iPSC derived enamel organoid guided by single-cell atlas of human tooth development. *Biorxiv.* 2022 Aug;2022-08.
- Xu X, Li Z, Ai X, Tang Y, Yang D, Dou L. Human three-dimensional dental pulp organoid model for toxicity screening of dental materials on dental pulp cells and tissue. *Int Endod J.* 2022 Jan;55(1):79-88.

22. Jeong SY, Lee S, Choi WH, Jee JH, Kim HR, Yoo J. Fabrication of dentin-pulp-like organoids using dental-pulp stem cells. *Cells*. 2020 Mar;9(3):642.
23. Chu J, Pieleś O, Pfeifer CG, Alt V, Morsczeck C, Docheva D. Dental follicle cell differentiation towards periodontal ligament-like tissue in a self-assembly three-dimensional organoid model. *Eur Cell Mater*. 2021 Jul;42:20-33.
24. Pringle S, Maimets M, van der Zwaag M, Stokman MA, van Gosliga D, Zwart E, Witjes MJ, de Haan G, van Os R, Coppes RP. Human Salivary Gland Stem Cells Functionally Restore Radiation Damaged Salivary Glands. *Stem Cells*. 2016 Mar;34(3):640-52.
25. Aihara E, Mahe MM, Schumacher MA, Matthis AL, Feng R, Ren W, et al. Characterization of stem/progenitor cell cycle using murine circumvallate papilla taste bud organoid. *Sci Rep*. 2015 Nov;5:17185.
26. Hisha H, Tanaka T, Kanno S, Tokuyama Y, Komai Y, Ohe S, et al. Establishment of a novel lingual organoid culture system: generation of organoids having mature keratinized epithelium from adult epithelial stem cells. *Sci Rep*. 2013 Nov;3:3224.
27. Driehuis E, Kolders S, Spelier S, Löhmußaar K, Willems SM, Devriese LA, et al. Oral mucosal organoids as a potential platform for personalized cancer therapy. *Cancer Discov*. 2019 Jul;9(7):852-71.
28. Karakasheva TA, Kijima T, Shimonosono M, Maekawa H, Sahu V, Gabre JT, et al. Generation and characterization of patient-derived head and neck, oral, and esophageal cancer organoids. *Curr Protoc Stem Cell Biol*. 2020 Jun;53(1):e109.
29. Shoushrah SH, Transfeld JL, Tonk CH, Büchner D, Witzleben S, Sieber MA, et al. Sinking Our teeth in getting dental stem cells to clinics for bone regeneration. *Int J Mol Sci*. 2021 Jun;22(12):6387.
30. Kim GH, Yang J, Jeon DH, Kim JH, Chae GY, Jang M, et al. Differentiation and establishment of dental epithelial-like stem cells derived from human ESCs and iPSCs. *Int J Mol Sci*. 2020 Jun;21(12):4384.
31. Hemeryck L, Lambrichts I, Bronckaers A, Vankelecom H. Establishing organoids from human tooth as a powerful tool toward mechanistic research and regenerative therapy. *J Vis Exp*. 2022 Apr;(182).
32. Li B, Ouchi T, Cao Y, Zhao Z, Men Y. Dental-Derived Mesenchymal Stem Cells: State of the Art. *Front Cell Dev Biol*. 2021 Jun;9:654559.
33. Chu J, Pieleś O, Pfeifer CG, Alt V, Morsczeck C, Docheva D. Dental follicle cell differentiation towards periodontal ligament-like tissue in a self-assembly three-dimensional organoid model. *Eur Cell Mater*. 2021 Jul;42:20-33.
34. Wang Y, Sun Y. Engineered organoids in oral and maxillofacial regeneration. *iScience*. 2022 Dec;26(1):105757.
35. Lassche G, van Engen-van Grunsven ACH, van Hooij O, Aalders TW, Am Weijers J, Cocco E, et al. Precision oncology using organoids of a secretory carcinoma of the salivary gland treated with TRK-inhibitors. *Oral Oncol*. 2023 Feb;137:106297.
36. Blutt SE, Estes MK. Organoid models for infectious disease. *Annu Rev Med*. 2022 Jan;73:167-82.
37. Chitturi Suryaprakash RT, Kujan O, Shearston K, Farah CS. Three-dimensional cell culture models to investigate oral carcinogenesis: A scoping review. *Int J Mol Sci*. 2020 Dec;21(24):9520.
38. Mohtasham N, Mohajer Tehran F, Abbaszadeh H. Head and neck cancer organoids as a promising tool for personalized cancer therapy: A literature review. *Health Sci Rep*. 2022 Apr;5(3):e580.
39. Lo YH, Karlsson K, Kuo CJ. Applications of organoids for cancer biology and precision medicine. *Nat Cancer*. 2020 Aug;1(8):761-73.